

Sample preparation of *Caenorhabditis elegans* (*C. elegans*) for RE-Pore-C extraction

24th July 2020

This protocol describes the preparation of a *C. elegans* sample to be processed with the restriction enzyme Pore-C (RE-Pore-C) protocol. The PD1074 strain of *C. elegans* was obtained from the Caenorhabditis genetics centre (CGC, which is funded by NIH Office of Research Infrastructure Programs (P40 OD010440). Worms were grown on solid agar media and fed with OP50 *Escherichia coli* (*E. coli*) over the course of 4–5 days at 21°C. The worms were predominantly harvested at the L1 larval stage following 1 day of starvation to reduce the presence of *E. coli* DNA in the worm gut. A worm pellet of approximately 2 ml worm pellet is used to yield approximately 5 ml of worm powder. This is sufficient for 5 preparations of Pore-C extract by following the RE-Pore-C protocol with approximately 1 ml of worm powder.

This protocol has been developed based on research by Oxford Nanopore Technologies and published literature: Jänes, J. et al. (2018) Chromatin accessibility dynamics across *C. elegans* development and ageing, *eLife*, **7**, pp. 1–24. doi: 10.7554/eLife.37344.

Materials

- Worms
- <u>M9 salt</u> solution supplemented with 1 mM <u>MgSO₄</u>
- Crushed ice
- Liquid nitrogen
- 15 or 50 ml centrifuge tubes
- Mortar and pestle
- Temperature-controlled 50 ml centrifuge
- -80°C freezer storage

Collection of worms: 20 minutes hands-on-time

Step 1

Gather sufficient worms by washing off the plate, to yield a pellet of approximately 2 ml following centrifugation in step 3.

Note: fewer worms may be used but reduced volumes will be required for subsequent washes and resuspension.

Step 2

Aspirate any supernatant above the worm pellet and then resuspend the \sim 2 ml worm pellet in 15 ml of M9 salt solution supplemented with 1 mM MgSO₄.

Step 3

Centrifuge at 800 g at 4°C for 5 minutes.

Step 4

Aspirate the supernatant and store the pellet on ice.

Preparation of frozen worm balls: 10 minutes hands-on-time

Note: Pre-cool the mortar and pestle at -80° C for at least 30 minutes.

Step 5

Resuspend the ~2 ml worm pellet in a minimal volume (~500 μ l) of M9 salt solution supplemented with 1 mM MgSO₄ final concentration, to achieve a worm slurry that can be pipetted.

Note: It is important not to over-dilute the worm slurry as this will decrease the ratio of worm:solvent and result in an underestimation of sample input for the subsequent RE-Pore-C extraction.

Step 6

Place the chilled mortar on ice and dispense a small volume of liquid nitrogen into the mortar.

Step 7

Using a 1000 μ l pipette tip, dispense the homogenous worm slurry dropwise into the liquid nitrogen to create worm balls of ~100 μ l.



Use a spatula to collect the frozen worm balls and store in a large chilled centrifuge tube on ice.

Step 9

Repeat steps 7–8 until all the worm slurry has been dispensed. Add more liquid nitrogen to the chilled mortar as required.



Snap freeze the tube of frozen worm balls in liquid nitrogen and store at -80 °C.

Cryogrinding of frozen worm balls: 10 minutes hands-on-time

Note: pre-cool a mortar and pestle at -80°C for at least 30 minutes.

Step 11

Place the chilled mortar and pestle on ice and add approximately 1 ml of frozen worm balls.



Carefully grind the balls into a fine powder, working quickly to minimise thawing.



Collect the worm powder into a chilled centrifuge tube on ice using a spatula.

RE-Pore-C extraction

Step 14

Transfer approximately 1 ml cryo-ground worm powder to a 50 ml centrifuge tube and resuspend in 1 ml chilled 1X PBS.



Bring the volume of the re-suspended cryo-ground tissue to 10 ml in chilled 1X PBS.

• Step 16

Proceed with the <u>RE-Pore-C protocol</u> using the re-suspended cryo-ground tissue powder as input.

Results

Sample	DNA concentration, ng/µl	Total DNA mass, µg
C. elegans	8.62	1.29

Table 1. The yield of non-size selected RE-Pore-C DNA extract using NlaIII restriction enzyme.

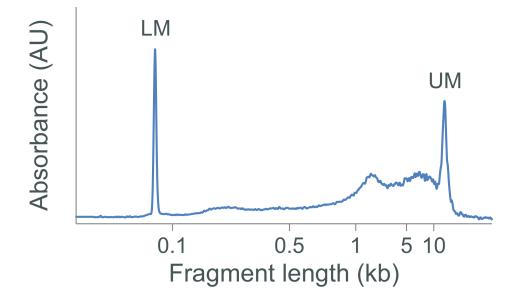
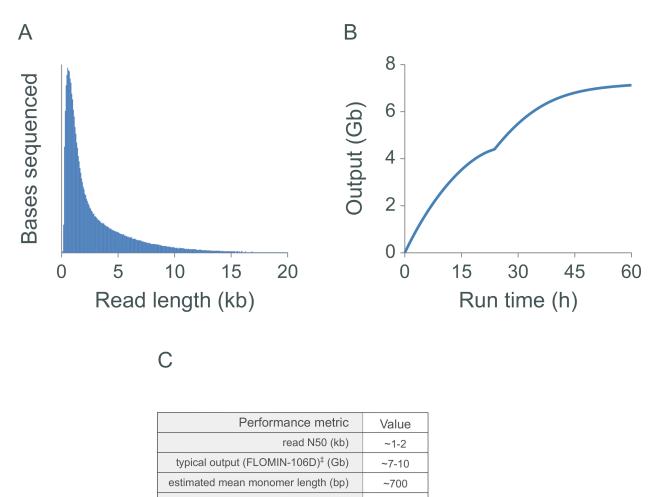


Figure 1. Agilent Bioanalyser DNA 12000 trace of non-size selected RE-Pore-C DNA extract.



~2	estimated monomers/N50 read	
~1 million	contacts/Gb	
~45	cis contacts (%)	

[‡] Nuclease flushes were performed to optimise flow cell output

Figure 2. The sequencing and Pore-C output for libraries assessed on GridION Mk1. Libraries were generated as described using Pore-C extracts prepared with NIaIII restriction enzyme. The read length distributions and output (Gbases) obtained from the library generated are shown in panels A and B, respectively. Panel C displays the Pore-C metrics obtained.

Date	Change note
24th July 2020	Updated formatting and added re-suspension of cryo-ground tissue powder.
5th November 2021	Included where the strain of C. elegans was obtained in the introduction.